

Ulva australis as a tool for monitoring zinc in the Derwent Estuary and implications for environmental assessment.

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Abstract

This study investigated temporal and spatial patterns of zinc content in Ulva australis. Samples were collected from the Derwent Estuary, Tasmania, Australia, between 2013-2015, at locations where zinc levels were elevated in both sediments and seawater historically. Zinc content was high (~321 mg·kg⁻¹) in U. australis at all sampling times, with levels consistent with the spatial distribution of metal within the system. Zinc in *Ulva* varied seasonally (5 - 321)mg kg⁻¹) and was highest in the middle-upper estuary, where seawater concentrations were highest, attributed to a nearby zinc smelter. The results suggest that spatial variation of zinc content in Ulva reflects the variability in the seawater, which in turn indicates that U. australis could be used for monitoring the effects of zinc in estuarine systems more broadly, and that U. australis could be a useful addition to existing management strategies in the Derwent and elsewhere.

Keywords: Biological indicators, Contamination, Macroalgae, Seasonal variability, Monitoring.

1. Introduction

Metal pollution in aquatic environments is a worldwide concern. Contamination of estuaries and coastal zones is increasing (Kennish 2002, Lionetto et al. 2012). Biological monitoring or biomonitoring is one way to obtain information on the potential effects of metals pollutants (Campbell 2002, Zhou et al. 2008). However, reliable indicators are needed to provide an understanding of the ecological effects (Rainbow 1995, Conti et al. 2002, Zhou et al. 2008). Measuring contaminant levels in seawater and sediments can provide a very accurate result (Campbell 2002), but does not provide a complete a complete picture of exposure or potential impacts from the contaminants. Measurement of levels in biota (flora and/or fauna) provides an better understanding of biological uptake over time, and as such may be a better indicator of overall ecosystem health (Holt and Miller 2011). Seaweeds have been recommended by a number of authors as potentially valuable bioindicators (Rainbow and Phillips 1993, Rainbow 1995). Many species have a high tolerance to metal pollution and being static will reflect local conditions (Zhou et al. 2008). However, there can be temporal differences associated with plant physiology (e.g. growth rate) (Malea and Haritonidis 1999b), environmental conditions (Malea et al. 1995, Brown et al. 1999) and even in pollutant inputs. The Derwent Estuary in Tasmania, Australia, is a highly metal polluted system (Bloom and Ayling 1977) with zinc (Zn) being a significant contaminant in the middle estuary (Coughanowr et al. 2015). Monitoring and research data collected over the past 15 years has shown that arsenic (As), cadmium (Cd) copper (Cu), lead (Pb) and Zn are all present at elevated concentrations in sediments (Bloom and Ayling 1977, Wood et al. 1992, Coughanowr et al. 2015), shellfish (Eustace 1974, Bloom and Ayling 1977) and fish (Eustace 1974, Langolis et al. 1987, Verdouw et al. 2010) in the Derwent. However, there is no information on the content of heavy metals in seaweeds in the Derwent. This study used Ulva australis, to characterise the spatial and temporal changes in zinc content within the estuary and examine the effectiveness of this species as a tool for monitoring.

2. Methods

2.1 Study area

Eight study sites were selected to reflect different metal loads and sources of pollution, i.e. levels of industrial discharge, sewage treatment levels, and heavy metal concentrations (Fig. 1). The estuary was divided into four regions where *U. australis* is located (upper, middle-upper, middle-lower and lower estuary).

2.2 Field collection

U. australis samples (n = 3) were collected from all study sites every three months from October (Spring) 2013 to October (Spring) 2015. Approximately 50 g of *U. australis* was collected from intertidal areas at low tide. Metal analysis was undertaken as per methods described by Farias *et al.* (2017). The Derwent Estuary Program (DEP) provided monthly data for surface water (0.1 m depth) zinc concentration (μ g·L⁻¹) for the middle estuary study sites (PWB: Princes of Wales Bay, SB: Shag Bay, KB: Kangaroo Bay, TRA: Tranmere, LSB: Lower Sandy Bay). All water and seaweed samples were analysed at Analytical Services Tasmania (AST).

2.3 Statistical analysis

Inter-annual differences were assessed between spring 2013 and spring 2015, using a general linear mixed model (GLMM) with area as random effect, year as factor, and gamma distribution. Where this showed a significant interaction, pairwise contrasts were undertaken using a Least-Square Means (Ismeans) package (Lenth and Hervé 2015), with Bonferroni adjustment. Seasonal variability was also analysed using GLMM, with area as random effect, year as factor, season as fixed effect, a gamma distribution and restricted suite of post-hoc multiple contrasts using the Least-Square Means (Ismeans) package (Lenth and Hervé 2015) and Bonferroni adjustment. Seasonal Zn variability in seawater was assessed using a GLMM with a gamma distribution. All GLMM analyses were performed in R with significance set at alpha = 0.05(R Core Team 2013).

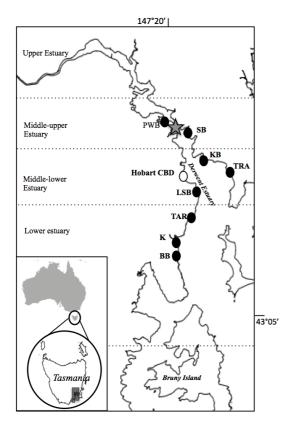


Figure 1. Study area and sampling sites in the Derwent Estuary, Tasmania, Australia. Grey star indicates Zinc smelter (historic source of metal contamination). Study sites were PWB: Princes of Wales Bay, SB: Shag Bay, KB: Kangaroo Bay, TRA: Tranmere, LSB: Lower Sandy Bay, TAR: Taroona, K: Kingston Beach and BB: Blackmans Bay.

3. Results

3.1 Inter-annual variability

Total Zn in *U. australis* content differed markedly between years; in 2013, mean Zn content was $320.7 \pm 18.3 \text{ mg} \cdot \text{kg}^{-1}$,

whereas in spring 2015 levels were considerably lower (Zn = $76.5 \pm 1.4 \text{ mg} \cdot \text{kg}^{-1}$) (Fig. 2).

3.2 Seasonal variability

Zn content varied seasonally (p <0.05, F = 8.076, df = 3) and spatially (p <0.05, F = 24.3439, df = 2). Seasonal changes were most pronounced in the middle-upper estuary (Table 1). Metal levels were relatively consistent over time, except in winter 2015, when Zn levels increased in the middle-upper estuary. The highest Zn levels occurred in the middle-upper estuary in spring 2013, and there was a clear spatial gradient in every season. There was a strong interaction between season and region (p <0.05, F = 4.0979, df = 6). Post-hoc analysis suggested that this was due to plants from the middle-upper estuary having a much higher Zn content $(244.3 \pm 108.0 \text{ mg} \cdot \text{kg}^{-1})$ compared to the middle-lower estuary $(42 \pm 27.2 \text{ mg} \cdot \text{kg}^{-1})$ and the lower estuary $(15.5 \pm 7.3 \text{ mg} \cdot \text{kg}^{-1})$ in all seasons (p <0.05, df = 5). This suggests that seasonal patterns are region-specific. Zn content was significantly higher in the middle-upper estuary in summer and spring, than in autumn or winter (Table 1). However, in autumn and winter, there were still clear spatial differences in Zn content in U. australis between the middle-upper and the lower estuary (p <0.05, df = 5). There was no significant difference in Zn levels between the middle-lower and the lower estuary (p > 0.05) in any season.

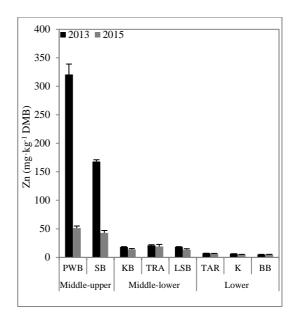


Figure 2. Inter-annual zinc (Zn) content (mg·kg⁻¹ DMB) in *Ulva australis* from sites in the Derwent Estuary in Spring 2013 and Spring 2015. Metal levels are expressed as mean (\pm SE). PWB: Princes of Wales Bay, SB: Shag Bay, KB: Kangaroo Bay, TRA: Tranmere, LSB: Lower Sandy Bay, TAR: Taroona, K: Kingston Beach and BB: Blackmans Bay. Note different scales on y-axis.

3.3 Metal content in seawater

There was significant spatial variability in Zn concentration in the surface waters (p < 0.05, F = 45.623, df = 1), with the middle-upper region of the estuary markedly higher than the middle-lower estuary (Table 1). This spatial difference was consistent over time, with no significant seasonal variability.

Table 1. Zinc (Zn) concentration in surface water ($\mu g \cdot L^{-1}$) and *Ulva australis* ($mg \cdot kg^{-1}$ DMB) collected from spring 2013, summer, autumn, winter and spring 2014 and 2015, from the three different regions defined in the current study. Metal levels expressed as mean (\pm SE), at Middle-upper estuary (n = 2), Middle-lower estuary (n = 3), and Lower estuary (n = 3).

Year	Season	Middle-upper		Middle-lower		Lower
		Water	Ulva	Water	Ulva	Ulva
2013	Spring	16.5 ± 1.5	244 ± 76.3	9.7 ± 0.3	19.3 ± 0.9	4.2 ± 1.3
2014	Summer	23 ± 1	156 ± 128.1	9 ± 0.3	24.8 ± 3.1	6.6 ± 1.1
	Autumn	26.8 ± 4.2	67.4 ± 41.6	9.4 ± 0.8	42 ± 15.7	15.5 ± 3.3
	Winter	28.8 ± 9.5	98 ± 39.1	12.6 ± 0.9	30.1 ± 1.2	8.9 ± 3.2
	Spring	20.5 ± 2.8	106 ± 35	13.2 ± 0.9	10.3 ± 1.3	4.2 ± 1.1
2015	Summer	22.5 ± 2.5	83.3 ± 55.8	14.9 ± 0.7	19.2 ± 3	6.5 ± 1.1
	Autumn	23.3 ± 2	50.6 ± 28.5	12 ± 0.8	17.2 ± 6.6	7.3 ± 1.1
	Winter	17.1 ± 1.8	73.2 ± 2.7	15.1 ± 0.2	35.2 ± 2.6	9.7 ± 3
	Spring	25.7 ± 0.7	47.2 ± 4.2	14.2 ± 0.4	15.5 ± 1.8	4.3 ± 0.9

4. Discussion

U. australis appears to be a good biological monitor for zinc in the Derwent estuary, broadly reflecting environmental conditions and providing a better understanding of the biological contamination. Ulva australis accumulated Zn showing a clear spatial gradient. The highest concentrations in algae were found in areas were the environmental loading was also high, i.e. middleupper estuary (PWB and SB), closest to the Nyrstar Zn smelter. The distribution of metal contamination in Ulva relates well to the patterns of contamination described by the DEP monitoring program (Coughanowr et al. 2015)Metal content in algae from the lower estuary was consistently low, and comparable to levels reported for unpolluted systems worldwide, suggesting that this region could act as a reference for the rest of the system. There were clear inter-annual and seasonal differences in zinc levels in Ulva throughout the estuary. This likely reflects a combination of both the scale of Zn contamination in the Derwent and the significant remediation efforts to capture industrial inputs over the last few years (Coughanowr et al. 2015). Zn is the predominant contaminant in the Derwent, and has been established as a suitable proxy for most other heavy metals contaminants (Cu, As and Pb) (Coughanowr et al. 2015). Zinc has been consistently detected at high concentrations in the water column, sediments, fish and shellfish (Coughanowr et al. 2015). Consequently, evaluating of Zn levels should provide a good indication of the broader metal contamination in this system. The temporal differences observed could be due to a number of factors. In a biomonitoring context, it is the differences in the metal loading/ inputs that are important, but temporal differences can also arise as a result of changes in the alga's ability to accumulate metals, that due to physiological status or its environment, both of which could be affected by the timing of sampling. In the current study, Zn levels in U. australis were highest in the most polluted (middle-upper) region of the estuary. In this area the contamination levels are very high and would likely obscure any subtle seasonal or temporal influences. Zn content in U. australis was greater in spring and summer than other times of the year, which is perhaps not surprising as this is the time when growth/ metabolic rate

was highest (in summer) and/ or when the plants were reproductive (in spring). Temporal variability in metal contamination in algae can be related to biological factors such as the growth strategy (annual/ perennial) or to the morphology of the particular species (Stengel et al. 2004). For Ulva, many studies have shown species specific differences: Ulva linza has been shown to take up metals to a greater extent in spring and summer (Haritonidis and Nikolaidis 1990), U. lactuca was found to have the highest Zn content in winter (Brown et al. 1999), U. rigida and U. intestinalis uptake was greatest in autumn/winter (Villares et al. (2002) and Haritonidis and Malea (1999) observed maximum metal content in U. rigida in autumn. These differences are likely due to changes in plant metabolism, which in turn will be affected by a range of environmental variables such as light and temperature. Previous research has shown that salinity can influence metal uptake in a range of Ulva species (Ho 1990, Malea and Haritonidis 1999a), with levels generally increasing as salinity declines (Villares et al. 2002, Mamboya et al. 2009). There is a reasonably strong salinity gradient in the Derwent estuary, with salinity dropping 10 ppt in surface waters through the middle estuary, the area most contaminated with Zn and where the highest levels were detected in U. australis. The levels observed in U. australis at these sites were clearly elevated, with biological implications for both the plants themselves and the food web. Should the salinity in this region decline (i.e. due to catchment management practices) this could affect the biomonitoring potential of Ulva in two important ways: i) directly - resulting in higher metal levels in the plants and associated increase in biological risk, and ii) indirectly - affecting the temporal and spatial comparability of results in a biomonitoring context. Consequently, it is important to consider salinity when using Ulva in a biomonitoring program. Using a biomonitoring species such as Ulva can provide an improved understanding of Zn contamination throughout the estuary over a biologically meaningful timeframe (the growing period of the plant), whilst samples from the water column will only ever provide a snapshot of the concentration at the time of sampling. The results showed that although U. australis broadly reflected a similar contamination gradient and pattern to that observed in the

water column/ sediment sampling, reflecting the different matrices. Monitoring U. australis could provide important information on changes in a bioavailable contamination source over time, something which has been missing in monitoring to date and which could provide an important new approach to evaluate the effect of remediation/ deterioration activities. Finally, U. australis has the added advantage of being both locally abundant and ubiquitous throughout the estuary. Previous studies have suggested that local species are generally the most suitable for biomonitoring (Phillips and Rainbow 1994, Rainbow 2006), and that cosmopolitan, widely distributed (O'Leary and Breen 1997) or abundant (Amado Filho et al. 1999) species are to be preferred. Currently seaweeds are not included in the Derwent Estuary monitoring program. Adopting U. australis as an indicator in this program, in conjunction with seawater and sediment monitoring, would provide another line of evidence with which to assess changes (improvement/ deterioration) in the "biological" condition of this metal-impacted system.

5. Conclusion

U. australis can play a valuable role as biomonitoring species, and can help improve our understanding of the potential risks and changes (improvements and deteriorations) in metal contaminated systems.

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References

- Amado Filho, G. M., L. R. Andrade, C. S. Karez, M. Farina, and W. C. Pfeiffer. 1999. Brown algae species as biomonitors of Zn and Cd at Sepetiba Bay, Rio de Janeiro, Brazil. Marine Environmental Research 48:212-224.
- Bloom, H. and G. M. Ayling. 1977. Heavy Metals in the Derwent Estuary. Environmental Geology 2:3-22.
- Brown, M. T., W. M. Hodgkinson, and C. L. Hurd. 1999. Spatial and temporal variations in the copper and zinc concentrations of two green seaweeds from Otago Habour, New Zealand. Marine Environmental Research 47:175-184.
- Campbell, I. C. 2002. Biological monitoring and assessment using invertebrates. Page 1100 *in* F. Burden, I. McKelvie, U. Fostner, and A. Guenther, editors. Environmental monitoring handbook. MacGraw-Hill Handbook Series, New York.
- Conti, M. E., M. B. Tudino, J. O. Muse, and G. Cecchetti. 2002. Biomonitoring of heavy metals and their species in the marine environment: the contribution of atomic absorption spectroscopy and inductively coupled plasma spectroscopy. Trends in Applied Spectroscopy 4:295-324.
- Coughanowr, C., S. Whitehead, J. Whitehead, L. Einoder, and U. Taylor. 2015. State of Derwent: a review of environmental data from 2009 to 2014. Derwent Estuary Program.
- Eustace, J. J. 1974. Zinc, cadmium, copper and manganese in species of finfish and shellfish Caught in the Derwent Estuary, Tasmania. Australian Journal of Marine and Freshwater Research 25:209-220.

- Farias, D. R., C. L. Hurd, R. S. Eriksen, C. Simioni, E. C. Schmidt, Z. L. Bouzon, and C. K. Macleod. 2017. In situ assessment of Ulva australis as a monitoring and management tool for metal pollution. Journal of Applied Phycology.
- Haritonidis, S. and P. Malea. 1999. Bioaccumulation of metals by the green alga *Ulva rigida* from Thermaikos Gulf, Greece. Environmental Pollution 104:365-372.
- Haritonidis, S. and G. Nikolaidis. 1990. Cd and Zn uptake in macrophyceae from Greek coasts. Biology Metals 2:235-238.
- Ho, Y. 1990. *Ulva lactuca* as bioindicator of metal contamination in intertidal waters in Hong Kong. Hydrobiologia 203:73-81.
- Holt, E. A. and S. W. Miller. 2011. Bioindicators: Using organism to measure environmental impacts. Nature Education Knowledge 3:1-10.
- Kennish, M. J. 2002. Environmental threats and environmental future of estuaries. Environmental Conservation 29:78-107.
- Langolis, D., R. J. Cooper, N. H. Clark, and D. A. Ratkowsky 1987. The effect off a mercury containment programme at a zinc smelting plant on the mercury content of sand flathead in the Derwent estuary. Marine Pollution Bulletin 18:67-70.
- Lenth, R. V. and M. Hervé. 2015. Ismeans: Leasy-Squares Means. R package version 2.16.
- Lionetto, M. G., R. Caricato, M. E. Giordano, E. Erroi, and T. Schettino. 2012. Carbonic anhydrase as pollution biomarker: an ancient enzyme with a new use. International Journal of Environmental Research of Public Health 9:3965-3977.
- Malea, P. and S. Haritonidis. 1999a. Metal content in *Enteromorpha linza* (Linnaeus) in Thermaikos Gulf (Greece). Hydrobiologia 394:103-112.
- Malea, P. and S. Haritonidis. 1999b. Seasonal accumulation of metals by red alga *Gracilaria verrucosa* (Huds.) Papens. from Thermaikos Gulf, Greece. Journal of Applied Phycology 11:503-509.
- Malea, P., S. Haritonidis, and T. Kevrekidis. 1995. Metal content of some green and brown seaweeds from Antikyra Gulf (Greece). Hydrobiologia 310:19-31.
- Mamboya, F., T. J. Lyimo, T. Landberg, and M. Björk. 2009. Influence of combined changes in salinity and copper modulation on growth and copper uptake in the tropical green macroalga *Ulva reticulata*. Estuarine, Coastal and Shelf Science 84:326-330.
- O'Leary, C. and J. Breen. 1997. Metal levels in seven species of mollusc and in seaweeds from the Shannon Estuary. Biology and Environment: Proceeding of the Royal Irish Academy 97B:121-132.
- Phillips, D. and P. Rainbow. 1994. Biomonitoring of trace aquatic contaminants. Chapman & Hall, London.
- R Core Team. 2013. R: A language and environment for statistical computing. R Foundation for Statistical Computing. Vienna, Austria.
- Rainbow, P. S. 1995. Biomonitoring of heavy metal availability in the marine environment. Marine Pollution Bulletin 31:183-192.
- Rainbow, P. S. 2006. Biomonitoring of trace metals in estuarine and marine environments. Australian Journal of Ecotoxicology 12:107-122.
- Rainbow, P. S. and D. J. H. Phillips. 1993. Cosmopolitan biomonitors of trace metals. Marine Pollution Bulletin 26:593-601.
- Stengel, D. B., A. Macken, L. Morrison, and N. Morley. 2004. Zinc concentrations in marine macroalgae and a lichen from western Ireland in relation to phylogenetic grouping, habitat and morphology. Marine Pollution Bulletin 48:902-909.

- Verdouw, J. J., C. K. Macleod, B. F. Nowak, and J. M. Lyle. 2010. Implications of Age, Size and Region on Mercury Contamination in Estuarine Fish Species. Water, Air, & Soil Pollution 214:297-306.
- Villares, R., X. Puente, and A. Carballeira. 2002. Seasonal variation and background levels of heavy metals in two green seaweeds. Environmental Pollution 119:79-90.
- Wood, J. M., P. Horwitz, and H. Cox. 1992. Levels of heavy metals in dated sediments from Lindisfarne Bay, Derwent Estuary, Tasmania. The Science of the total environment 125:253-262.
- Zhou, Q., J. Zhang, J. Fu, J. Shi, and G. Jiang. 2008. Biomonitoring: An appealing tool for assessment of metal pollution in the aquatic ecosystem. Analytica Chimica Acta 606:135-150.