

Short-term effects of elevated temperature and CO₂ on *Pisum sativum*

Juozapaitienė G.¹, Dikšaitytė A.¹

¹ Department of Environmental sciences, Faculty of Natural sciences, Vytautas Magnus University, Vileikos St. 8, Kaunas, Lithuania.

*corresponding author: Juozapaitienė G.

e-mail: g.juozapaitiene@gmf.vdu.lt

Abstract The concentration of CO₂ is predicted to further increase till the end of this century and the changes in global temperature are also expected. Due to the fact that increasing CO₂ and temperature will change the agricultural environment we investigated photosynthesis, plant productivity and organic carbon (C_{org}) accumulation of pea (*Pisum sativum* L.) under increasing levels of air temperature and atmospheric CO₂. A closed growth chamber experiment was performed in a controlled environment at ambient [21 °C/400 ppm] and elevated [25 °C/800 ppm] temperature and CO₂ conditions.

The results showed that after 4 weeks of treatment aboveground (49 %, p<0.05) and belowground (6 %, p>0.05) biomass of pea increased, also significantly increased photosynthesis (54 %) and leaf area (37 %) under elevated climate [25 °C/800 ppm] conditions compared to the conditions of [21 °C/400 ppm]. Either it was estimated that organic carbon partitioning in leaves and stems increased (p>0.05) under elevated climate conditions [25 °C/800 ppm], but decreased in roots (p<0.05).

Our results suggest that the effects of increasing levels of air temperature and atmospheric CO₂ were greater for photosynthesis, leaf area and aboveground biomass of pea. Also results demonstrated a promising potential in biomass C accumulation.

Keywords: climate change, photosynthesis, organic carbon, closed chamber experiment, *Pisum sativum*

1. Introduction

It is expected that CO₂ value could increase to an atmospheric concentration of between 750 and 1300 ppm for the end of the century and temperature is predicted to increase between 1.8 and 6.0 °C (IPCC, 2014).

Elevated CO₂ concentrations directly influence soil-plant systems via enhancing plant growth (Butterly, 2015). Elevated global temperature in the future will impact ecology and agriculture and may prove to be a major factor limiting crop production (Kurek *et al.*, 2007; Perez *et al.*, 2009; Qin *et al.*, 2008). Climatic changes due to

temperature, water availability variation, and increase in CO₂ will also affect the carbon–nitrogen balance in the plants and eventually how seeds set and grow (Vadez *et al.*, 2012).

Since global increases in temperature and CO₂ may have interactive effects on photosynthesis, many studies have examined the effects of elevated CO₂ and increased growth temperature (typically 3–5 °C) on photosynthesis (Morison and Lawlor 1999). Only a few studies have examined the interactions between elevated CO₂ and higher mean growth temperature on plant carbon accumulation. According to Dyson (2008) “If we can control what the plants do with carbon, the fate of the carbon in the atmosphere is in our hands”. That is why one of the aim of our research is to analyze carbon partitioning in legumes plants. Legumes provide important sources of oil, fiber, and protein-rich food and feed while supplying nitrogen (N) to agro-ecosystems via their unique ability to fix atmospheric N₂ in symbiosis with the soil bacteria rhizobia, increasing soil carbon content, and stimulating the productivity of the crops that follow (Jensen *et al.*, 2011)

2. Methods

The experiment was conducted in a closed controlled environment plant growth chambers located at Vytautas Magnus University in 2016. Seeds of pea (*Pisum sativum* L., cv. Pinochis) (15 individuals per pot) were planted in 3 L plastic pots (diameter 21 cm, height 10.6 cm) containing a growth substrate composed of a mixture of field soil (the soil was taken from ASU Training Farm, Kaunas District), perlite and fine sand (5:3:2, by volume). A nutrient supply corresponding to 120 kg N ha⁻¹ was used until the beginning of treatment. Additional fertilization with a complex nutrient (NPK 12-11-18 + microelements) solution, increasing the N level until 180 kg N ha⁻¹, was applied two weeks after the treatments.

Elevated CO₂ and temperature (day/night temperature of 25/18 °C and 800 ppm of CO₂) treatment was applied when the seedlings of pea were germinated and lasted for 4 weeks. Until that time all plants were grown in the control chamber under conditions of current climate – an average day/night temperature of 21/14 °C and 400 μmol mol⁻¹ of CO₂. The following stable conditions were maintained in

both chambers: a photoperiod of 14 h, relative air humidity of 60–70%, and $200 \mu\text{mol m}^{-2} \text{s}^{-1}$ photon flux density of photosynthetically active radiation. The chamber was 2.5 m high, 2 m wide and 2 m long. The pots in the chamber were watered sufficiently and regularly. All treatments were run in three replicates.

Gas exchange was measured with portable photosynthesis system LI-6400 (LI-COR, USA) with randomly selected youngest fully expanded leaves at the end of experiment. The photosynthetic rate determined at the corresponding growth CO_2 concentrations (A_{growth} , $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$), stomatal conductance ($g_{s(\text{growth})}$, $\text{mmol H}_2\text{O m}^{-2} \text{ s}^{-1}$), transpiration rate (E , $\text{mmol H}_2\text{O m}^{-2} \text{ s}^{-1}$) were recorded automatically for approximately 5 minutes.

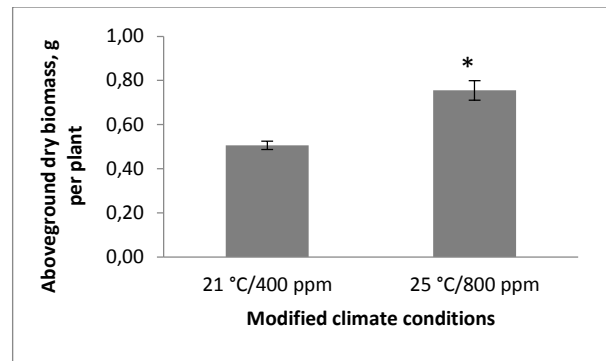
The measurement of leaf area was carried out on the last day of treatment. The leaf areas of all leaves per plant of five plants per treatment were measured with a scanner (CanoScan 4400F, Canon, USA) and then the leaf area of all leaves per plant was determined by GIMP 2.8 software.

For determination of dry weight, plants from each pot were harvested and the samples of leaves, stems and roots were dried in an electric oven at $70 \text{ }^\circ\text{C}$ until a constant dry weight was obtained. Biomasses of the samples were calculated into dry biomass per plant. Measurements of carbon accumulation were carried out at a 28-day period after the treatment. A subsample of plant roots was dried in an electric air-forced oven at $70 \text{ }^\circ\text{C}$ until a constant dry weight was obtained (at least 72 hours). Soil samples were also taken at a 28-day period after the treatment. The samples were air dried at room temperature and sieved through 2 mm mesh on purpose to remove all visible roots and plant remains. The dried samples of shoots, roots and soil were ground to a fine powder with a mill (Retsch HM400, Germany). Organic carbon content was measured with a Shimadzu TOC-V solid sample module SSM-5000A in the laboratory of Vytautas Magnus University.

The independent-samples t-test was applied to estimate the difference between reference and treatment values and p value < 0.05 was the threshold for significance. All analyses were performed by STATISTICA and the results were expressed as the mean values and their standard errors ($\pm\text{SE}$).

3. Results and discussion

The results showed that after 4 weeks of treatment aboveground biomass of pea (Figure 1) significantly ($p < 0.05$) increased 49 % under elevated climate conditions [$25 \text{ }^\circ\text{C}/800 \text{ ppm}$] compared to ambient climate [$21 \text{ }^\circ\text{C}/400 \text{ ppm}$] still there was only a small increase in belowground biomass of pea (Figure 2) (6 %, $p > 0.05$) under elevated conditions, compared to ambient.



Note. * – statistically significant difference at $p < 0.05$, as compared to the reference treatment [$21 \text{ }^\circ\text{C}/400 \text{ ppm}$]

Figure 1. Dry aboveground biomass of pea at different climate conditions (mean \pm SE)

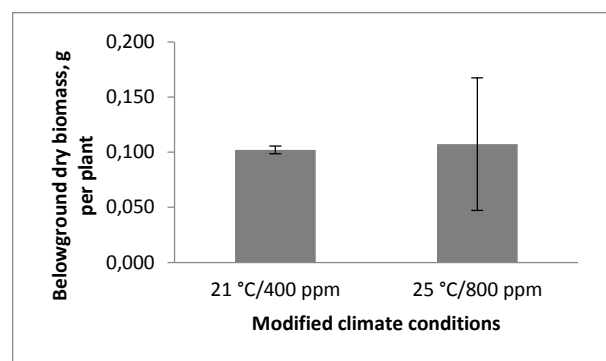
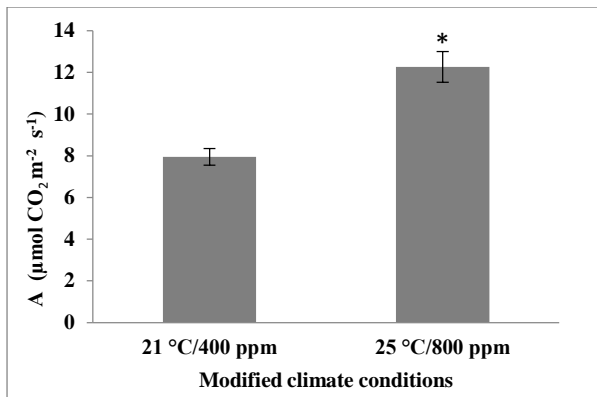


Figure 2. Dry belowground biomass of pea at different climate conditions (mean \pm SE)

According to some authors (Erice *et al.*, 2006; Irigoyen *et al.*, 2014) plants usually grew more when exposed to a combination of elevated CO_2 concentrations and elevated temperature than under current CO_2 and temperature conditions. However Juknys (2011) investigated, that increasing biomass of pea under elevated both CO_2 and temperature were positive but not statistically significant. The change of aboveground biomass in our treatment was bigger compared with other authors results. For example, Butterly (2015) found that elevated CO_2 increased shoot biomass of field pea by 36 %.

Increasing levels of air temperature and atmospheric CO_2 significantly increased photosynthesis (54 %) (Figure 3). According to Irigoyen (2014) when the atmospheric CO_2 concentration rises suddenly (or in a temporal window up to a few days) from 400 to $700 \mu\text{mol mol}^{-1}$, the photosynthetic C fixation of C_3 plants increases. Either an increase in photosynthesis by 25 – 75 % has been detected in many experimental studies on the impact of doubled CO_2 concentration on C_3 crops (Urban, 2003; Kirschbaum, 2004).



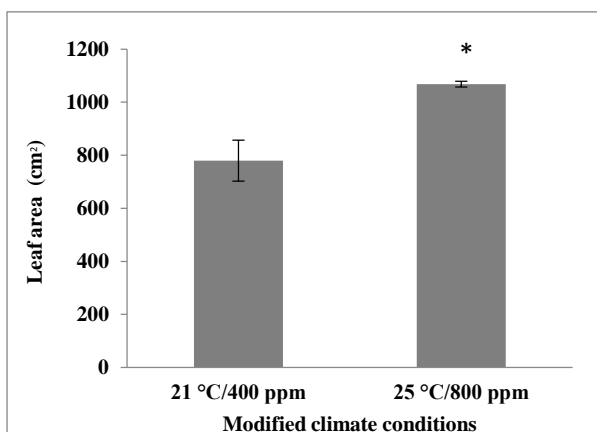
Note. * – statistically significant difference at $p < 0.05$, as compared to the reference treatment [21°C/400 ppm]

Figure 3. The rate of photosynthesis under different climate conditions

The effect of changing climate on transpiration and stomata conductance was not statistically significant ($p > 0.05$) with a change accordingly to 2 % and -4 %.

Reduced stomatal conductance in a higher CO₂ environment will maintain plant water relations, but may have implications for heat stress as leaf temperature rises with reduced transpiration (Vadez *et al.*, 2012).

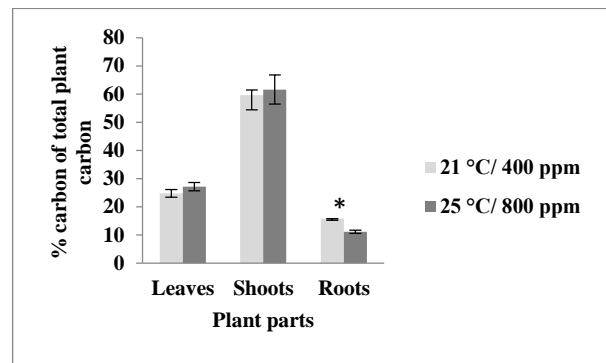
It is said, that a prevailing response of crops to rising CO₂ is an increase in leaf area (Srinivasan *et al.*, 2016). According to our results, leaf area significantly increased 37 % under elevated climate conditions [25 °C/800 ppm] compared to ambient climate [21 °C/400 ppm] (Figure 4). For example, model simulations showed that soybean crops grown under current and elevated (550 ppm) CO₂ overinvest in leaves, and this is predicted to decrease productivity and seed yield 8% and 10%, respectively (Srinivasan *et al.*, 2016).



Note. * – statistically significant difference at $p < 0.05$, as compared to the reference treatment [21°C/400 ppm]

Figure 4. Leaf area under different climate conditions (mean ± SE)

Organic carbon partitioning in leaves and stems increased ($p > 0.05$) under elevated climate conditions [25 °C/800 ppm], but decreased in roots ($p < 0.05$) (Figure 5).



Note. * – statistically significant difference at $p < 0.05$, as compared to the reference treatment [21°C/400 ppm]

Figure 5. % carbon of total plant carbon in plant parts under different climate conditions

Decreasing carbon in roots can be explained as nodules are competing for carbon use with the roots (Voisin *et al.* 2003a). As such, nodulation can limit root growth (according to our results, there was only 6 % increase of pea roots biomass (Figure 2) ($p > 0.05$) under elevated conditions, compared to ambient). When net photosynthesis rate per plant increased, net sink strength also increased, suggesting that the C supply from photosynthesis was limiting. It has been shown by our treatment and other authors (Voisin *et al.*, 2003b) that the percentage of C respired by belowground parts decreased when net photosynthesis rate increased. This suggests that the amount of C respired (for synthesis, maintenance of structures and/or symbiotic N₂ fixation or root mineral N assimilation activities) was decreasing the C supply available for investment in biomass of the root system (Warembourg, 1983) (Voisin *et al.*, 2003b).

4. Conclusion

The results presented here demonstrated that, elevated temperature and CO₂ conditions positively affected dry aboveground and belowground biomass of pea; photosynthesis and leaf area also increased under [25 °C/800 ppm] climate conditions. Although organic carbon partitioning in leaves and stems increased under elevated climate conditions [25 °C/800 ppm], but decreased in roots because of nodules were competing for carbon use with the roots.

Further researches are needed to investigate the alteration of C/N ratio in legumes under elevated climate conditions

References

- Butterly C. R., Armstrong R., Chen D., Tang C. (2015), Carbon and nitrogen partitioning of wheat and field pea grown with two nitrogen levels under elevated CO₂, *Plant Soil*, **391**, 367–382.
- Dyson F. (2008), *The Question of Global Warming*. New York Review of Books. (30 June 2010);

- Ericc G., Irigoyen J. J., Pérez P., Martínez-Carrasco R., Sánchez-Díaz M. (2006), Effect of elevated CO₂, temperature and drought on dry matter partitioning and photosynthesis before and after cutting of nodulated alfalfa, *Plant Science*, **170**, 1059–1067.
- IPCC. (2014), Summary for policymakers. In: *Climate Change 2014: Impacts, Adaptation, and Vulnerability. Part A: Global and Sectoral Aspects*. Contribution of Working Group II to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. Eds. C. B. Field, V. R. Barros, D. J. Dokken, K. J. Mach, M. D. Mastrandrea, T. E. Bilir, M. Chatterjee, K. L. Ebi, Y. O. Estrada, R. C. Genova, B. Girma, E. S. Kissel, A. N. Levy, S. MacCracken, P. R. Mastrandrea, and L. L. White. Cambridge, United Kingdom: Cambridge University Press. P. 1–32.
- Irigoyena J. J., Goicoechea N., Antolína M. C., Pascual I., Sánchez-Díaz M., Aguirreolea J., Morales F. (2014), Growth, photosynthetic acclimation and yield quality in legumes under climate change simulations: An updated survey, *Plant Science*, **226**, 22–29.
- Jensen E. S., Peoples M. B., Boddey R. M., Gresshoff P. M., Hauggaard-Nielsen H., Alves B. J. R., Morrison M. J. (2012), Legumes for mitigation of climate change and the provision of feedstock for biofuels and biorefineries. A review, *Agronomy for Sustainable Development*, **32**, 329–364
- Juknys R., Duchovskis P., Sliesaravičius A., Šlepetyš J., Januškaitienė I., Brazaitytė A., Ramaškevičienė A., Lazauskas S., Dėdelienė K., Sakalauskaitė J., Juozaitytė R., Kadžiulienė Ž., Dikšaitytė A. (2011), Response of different agricultural plants to elevated CO₂ and air temperature, *Agriculture*, **98**, 259–266.
- Kirschbaum M. U. F. (2004), Direct and indirect climate change effects on photosynthesis and transpiration, *Plant Biology*, **6**, 242–253.
- Kurek I., Chang T. K., Bertain S. M., Madrigal A., Liu L., Lassner M. W., Genhai Zhu G. (2007), Enhanced thermostability of *Arabidopsis* Rubisco activase improves photosynthesis and growth rates under moderate heat stress, *Plant Cell*, **19**, 3230–3241.
- Morison J. I. L., Lawlor D. W. (1999), Interactions between increasing CO₂ concentration and temperature on plant growth, *Plant Cell Environment*, **22**, 659–682.
- Perez D. E., Hoyer J. S., Johnson A. I., Moody Z. R., Lopez J., Kaplinsky N. J. (2009), BOBBER1 is a noncanonical *Arabidopsis* small heat shock protein required for both development and thermotolerance, *Plant Physiology*, **151**, 241–252.
- Qin D., Wu H., Peng H., Yao Y., Ni Z., Li Z., Zhou C. Sun Q. (2008), Heat stress-responsive transcriptome analysis in heat susceptible and tolerant wheat (*Triticum aestivum* L.) by using Wheat Genome ArrayBMC, *Genomics*, **9**, 432.
- Srinivasan V., Kumar P., Long S. P. (2016), Decreasing, not increasing, leaf area will raise crop yields under global atmospheric change, *Global Change Biology*, **23**, 1626–1635.
- Urban O. (2003), Physiological impacts of elevated CO₂ concentration ranging from molecular to whole plant responses, *Photosynthetica*, **41**, 9–20.
- Vadez V., Berger J. D., Warkentin T., Asseng S., Ratnakumar P., Poorna Chandra Rao K., Gaur P. M., Munier-Jolain N., Larmure A., Voisin A., Sharma H. C., Pande S., Sharma M., Krishnamurthy L., Zaman M. A. (2012), Adaptation of grain legumes to climate change: a review, *Agronomy for Sustainable Development*, **32**, 31–44.
- Voisin A. S., Salon C., Jeudy C., Warembourg F. R. (2003a), Root and nodule growth in *Pisum sativum* L. in relation to photosynthesis. Analysis using ¹³C labelling, *Annals of Botany*, **92**, 557–563.
- Voisin A. S., Salon C., Jeudy C., Warembourg F. R. (2003b), Seasonal patterns of ¹³C partitioning between shoot and nodulated roots in *Pisum sativum* L. under different air CO₂ concentrations, *Annals of Botany*, **91**, 539–546.
- Warembourg F. R. (1983), Estimating the true cost of dinitrogen fixation by nodulated plants in undisturbed conditions, *Canadian Journal of Microbiology*, **29**, 930–937.